

BIOINFORMATICS WORKSHOP

Sometimes genetic mutations in an organism can lead to it being more or less susceptible to certain drug treatments. This is particularly important when we are looking at the effectiveness of drug treatment against parasitic worms as there may be resistant and susceptible strains of worms. In this workshop we will be using an example sequence to design primers for and then we will analyse. To do this we will be using bioinformatics. In this workshop we are aiming to do the following:

1. Understand how the primers are designed for the PCR process.
2. Look for mutations in our sequence to identify our resistant and susceptible strains.

When we are designing primers for our PCR we need to think about a few things in order to get a good primer designed for our sequence. Good primer design is essential for a successful PCR. In order to design your primer there are some important considerations, which include but are not exclusive to the following:

- a. Primer length
- b. Primer melting temperature
- c. Primer annealing temperature
- d. GC content

1. **Primer length** – Optimal primer length is generally considered to be between 18 to 22bp long. This is long enough for the primer to anneal to our DNA easily and to be specific for our DNA region.
2. **Primer Melting Temperature (T_m)** – This is the temperature where the double stranded DNA dissociates to become single stranded DNA. Temperatures generally range between 52 and 58°C. Secondary annealing can occur with temperatures above 65°C.
3. **Primer Annealing Temperature (T_a)** – This is the temperature required for the primer to anneal to the DNA sequence. Too low a T_a and the primers may anneal to sequences that we are not targeting therefore leading to a reduction in the product yield. Too high a T_a may also lead to a low product yield as the primer annealing may be reduced.
4. **GC content** – This is the number of G's and C's in the DNA sequence. The total of which should be between 40 – 60%.

We will design primers using a primer design software tool called PRIMER 3 (Rozen and Skaletsky, 2000) for the example sequence below:

```
TTTCCGACGACGACGACGACGAAAGGGCAGCTTCGACATCGTCTGACGAAATACGGTAGTGACCTCAGAAAAGTCAGAATACAAGTAGTAGCAATAAGGTACAGTAGCAAGAG
TTGTGCAGTGCAGTTTCTTAGGGAGAGCAGAGCAGATAAAAATGGTGAATCGACACTGTGAGCCATCATGCGAGTGTATAGGCACTAAGTTTGTATCAGATAGTATCATTT
TGAAAAACAAGATTGTGCATTAGGACATATATAGTGGCAAAATGAGGTAACACAGCCAAAAGAGCGGTTAAATTTGGACTATGCGATCCCCATCTTTATTGGGAAGAGAAAA
AATACAAAAGGATTAACAACAAAATGCTTGGATTACAACACAGCAAGGGAAGAAAGAAATCTGGCAGAACAGGAATGGCATTGTTCAAACCTCCATCTCAATTTGTCCTGC
CATCCAAACGTTTATGTCATGCCATTAGATAAACCTCCAAACTCTTTGGCGCTATACTAGCTATTTATCCCGCTGCAAAAGATGTCTGCAGAAATAGCCGAAGGAAGGTAATTTCC
ACCACCAGGGGAGCCAGAAATATAGGAAGATTTGAAGAGAAAAACGAAAGAGGAGAGGGTACACTCAAGAGTTTCACTCCATCAGGCAAGTATCAAAAAGACTAGTCGACCT
CCGATACTGGGATGCGAGGCCGCAAGGAGAGCGGCTCCGATGCCAGACTCTCGTTGGCCAGCTTAGCAACCAGCGCAGAAAGAGGCTTCTCCCGAGGAGATCTGTA
AGAGTTGCTTCTAAGCAACTGCTGAACCTTTTGTAAATGTTTCATAGAGCCACCTCCAAGCAACGACCGTCTTGGCATCGTGTGCCATCAGTTGGCATCTTTGTCACGGC
CTAGCTTCTCAGGATGCCATATATACCAGCAGCGGCCAAGCGTGTCTCCGCGCTCAGCAACAATGTCACATATGTGACGAGTAATGTATCTTACTCCAGGGAAGTATCTGCA
ACCTGAACATGAAATCGAGATGTATACAAAGTAATCAATGGTCAATGAACATTTCTGGTATCCAGTTAGGTTACGCAAGTCAATACCCCAACGATCTTTAGCTTAGTTCCTC
AGAACTTTAAGGTCATGTGATGCATCATGGTGCATGGCTGACATATCGGGGCTCGCAATGCATAAGATTCAGTAGTTGAATTTATACGGAGTAAGAAATACACTACAACAACT
TCATACAGATAAACGCTTTAATCGTAATTTGAGCTCATGTTTCAGAAAACATTTGTCAGAAATTTAACAACAGATTTCTGCTACTAATGGCATGGGCTTCTCATAGTTCTCAAGAAAT
TCTATGCTATTCAATACACATATCAATCAACAATATAGTATATCTACTTAGCCTCAAGTAAATTTTCGATTAGTTCCAAGAAAGAGGACATAACACTATTCCACATCACTTTTG
AGCCACTTGAAAAACAAGAAAAAGGAAAAACAACACTTTGGTGTAAATGAAATTAAGGTTGTCAGCAACAACTCTATCCAAGTAAGCAATTTGCACAGCCCAT
CAACCAATTTATCTAATACCCAGAAAAAAGATACCAAGCTTTGAATTTGATCCGAGTTGAGGACATAACAGCAAAAACAGAGCAATTTGACCGATGTAAGATTTTCCACCGAT
GATGAATCGGCATTAATTTGATTTTGGCCCTCACATAGTACTTACTGAGTAATGTTAACTTCAGAAAGCCGAAATTTGCCAATCTTTTATAGCTTTGGGCTTTG
GCCTTACACGGACTAGTCAACTGATGTAACATGATAAATTCGGACGAGAATCTGTTTCTTGGCGAGTTTGAAGAGCTCAATATAGTGTGTAATAAATTTGATCACCCTTT
AGCTACAGCAAAAGATGAAGAGTTCAATGACTAATGGGTCAAGAAAGTATACCTCAGTCAAAACGGCTGTTCCAACCTTAGGTGGAACAACATCTCCAAACAATGCAGCATCA
TGAGCCAGTTTCAGTAAGATTTCTTCGCACAATCTCTCAAGATACATGCCAGAAATCACTTTTCATATATCTGTTTACCCAGAAAGATATTCCGATATATAGTCAGTTTAGCAA
CAAAACATAAATTTGGAAGTGAATAATACCACCGAGATAAACACCTGTTCTCCAGGATTCAGACTTTCAAAAATCCATAGCTTTATCGTATTTGAAAGTGAAGCTTTGCTGATTT
AAAGCTTCCCTTCCGTTGATACCTAGAGAACATCAACAATTTGATTTAGGTTAGGATGATGACAGCAAGTGAAGATACGATGTCACGCCCATACATGATGTTA
ATGGGACTTGTTCATTAACAGCAGAAAGGAAATTTAAACACCAACCATATTTCCGGATTTAGGCAAGTAACTCCAGTCCATTTTGGAAATTTGAAATGATTTAATAGCCACAG
GCATTTGTCAGTGCACAAAATACAGCAGCAACAACATCGTTATCCACATATCTCCACCAGCAATGTGCCGACTGTGTCATTAACCTACAAAATATCAACAATCAGCCT
ACATAAAGGAAATCTCAAAAAGAAAGAAAGTTCTGCATGACGCAAGCATGTTTCCATCCCTCAACAGAAAAGATTAACGAGTAGCATCAGGACACACAAAATAAA
AATTTGCATTTATTTGGGGAGTAAAAAATAGATGTTGTTAATACCAATGCTGTTAATACCAATGCTTCTCATATCCAAACCTCGCTTCCATCCACTCAGCCACAACTCCTC
ACCAACCTGATTAGGGAGTAACCAAAATAACAATTAATAAATCAAAAGTAAATTTGGTATTATGGGATGATTGAAAAATTTGGCTCAGAGACCATCAACCGAGAAAATTAATAT
CTTAAGTATCAAAAATATTTCTTCTTACCAAGCTTACATTTTCACTGTTGATGATAAATCAGATAAATCAACCCAGAAACATACTACTTACTGCTCATAAAAAATGTCCT
TCCTAACCAAGCTTTCATTTTATCATATTGAGATAAAGAAATCAGTTTGGACATAATGTAATGATGATAACGAAAGTATCAGTGAACATAAATCAACATGAAATAATTTA
ATAACATTTACTATTGAGATAAAGAAATCAGTTTGGACATAATGCAATGACTGATTTGAAAGTAACTAGAGCAAAATGAAATAATTTAATAGCCACAG
CCCACCACTATTAAGTTCAACACTTTGGCATATACAAATATACAAATGTTTCAGAAAGGCAAACTCTTTTAGCCCTCATGCCATCAGAAAATGAAATCATCTCTATT
ATATAGTTAACTAAATTAATAATTTGAAAGTGAATGACGAAAGATGACAATAGTACAATGAAAAAATGTCATAAGTGTCAAGAGGCAAACTCTTTTTCATTTCTCTCTC
AAGGTCAGACATTTATTTAGATGGTTTGGCTAAGAAAAGGGTGTGTTAAATCTCTCTACTATACCAAAAGAGTGATCAAGGACAAGAACCGGATCCCTTGCATATCAAAAT
AAAGGTTAACCCAGAAACTTTGCCATATCAATAAAAATACACTTGAAGCCATGTCAACTAATGTCTAGGAGTTTGTGGACATAATTCGATAGTGTGTTGAAATTTACCGT
GCCGTTGATCGAAAAACCTTTTCCACTTGTATGAGCGTTCCAGATGATATCGACAATTTGGTTCCTGAAAGGAAAAGGTAACCCAGCTCTCTGCTCCCTCGG
GAGGTTGAAATCGTTGCTTCCGTTATCCAAAATTTTGGCAATGCAGCAGCAATGAAATCAAAAGTTCTGTATCAATAGTTTAAAGATACATTAGGAACCCAGCGGCT
AAATTTGGCAACTCCAGAACTTGAGAACTCAACAGCAGGAACTCACCATGGAAGTCCCAACCTAGGTTGAGGTTGGATAGAGACTCTCCGACTGTGCTGGCA
ACACGTTTCTCCCTTCCGCGAGTTGGACTCGGAGTACACGGAAGTTGGTTCTCAAAAGTCCAGCCGCAAAAACAGCCCATGCTCGTCTGCAGACATAGCACCCAGAT
GATTAACACAAAAGTTCAAGAGTCAAGAAAAAATGCAATGTTTATCAGTGAATGCAACATAATAGTCTGAACGCAACAAAATGTTAGATTTAAGAACTTTGAGTTTCACT
GAATGATGCTCAACATGTTAAAGCAATTTATATACATCTGTTTATAAATTCGTGACCAATGTTTCTCAACATGAGTTCGATCTGAAAATTTCTAGTGTATAAGAGGATGATCCTAG
AAAACTAAGCAATTCACAGTTATACGATAGGTCGAGTAATGATATAACTAATAGAGAAATTTGCTGACACAAAACATAAAGATGATCAACGTTTCTAAGAAATTAAGA
GAATTTGAGCTTATAGGAGTAAATCTGAGATAACTACGATGTCATTAATCAAAAACCTAAGCACTTTGACCGATTATATCTAGAGATGATTTAGGTTGACTGGCAGATATACTG
AACAAAACAATTTGAAAGAAACGATAAAGACATATGAACTTTCTCAGAAATTCAGACTAGGTTGTAACATGCGCTAACTATCAATGCCATTAATTTGGACAGTACATGT
ACATCTCACTAAGGGCTATCAATAGGCTCGCCACCAGGCACTCATCCGAATCTCATATGCTCACTCCAAAGACTGGTGACATTTCTATTGACACCAATATCTAAATCATC
AAGTTGGTTTGGAACTACACGTTGATCTGCAAGCATGGGATTTGGCAGGGGCAAAATGTTGATGTTAAAGGTAAGAAATCTTCTGGCCATGATCCATGAAATCGGCAAGG
TCTGCATTTCCGCAATAAGATCCGGGCTTTCCGCGGAGGGCGGCTAATCTCAATCAAGAGCGGGAGAGAAAGAAATGCTGACGCTTCCGCGATCAGGACTCAGAAATCG
TCCAGCGCGGACGACAGTCCCAACAACAGACGGCGTCTTTGGGATCACAAACAAGTCAACCGAACCGCAAAAAGGAAAGGAAAGGAAAAAAGGAAAGGACCGG
GATCGGATCGGATCAGATCGGACAGAAATGGGATCTTTGTTACCCGGTGGGGAGGTTGTCGACATGATGAGATGAGCATCTTGAAGTGTGAGTGGATGTCGCCCGCAGTCC
CGCTTCCATCTCGCGCAGAGCGGCTCGGAGATGCTCCTGAGCAGGGCGTGGGCGTGGAGAGCGCGGCTCGACCTCCTCGATACGCTCCGCCACTTTGCGCTTCCGCT
CGGCTCGGCGGCGGCTGACGCTCCGCGCCCGCTTCTGGCGGCGCGGCCAGCAGCAGCCCGTGGCCACCGCGGCAACCAACCCCGCTGCCCCGCGCCG
CGCTTCCCATCGTGTAGATCAGCTTCTCCGCCAGCGAAAGGCGATCGACCGGCAAGAAATCGCAGAGGAATTCGCTCTGGATCCGAGGAGTGAAGGAAGGGAT
GGTTCCGCGGCGGCTAGGAGAGGGACTGGCCCTCGGCGTTTAAACGGTAACCTTTAACTGGCCCTTGGGATTTGCGGCAATTTATGGATCCACTATAAATACAGCAAGCGG
CGGATTTGTCGGGGGACACTGTCAGCAGCGGGATCTGGCCCTGGGAGAAAGTGGATCTGGATTTTTTGGATTTATAGGGAACGGATTTCTGCTGTGGAATATTACTAG
ACGGATTTACTCCAGATATACTTACGGTAAGTACTCCCGTGTAAATTAACCTAGGAGTATGATTTGGCGAGTAGATCTGAAATAGAGCGCTGACTTTGGCGGCGCC
CGGCTCCAGTGG
```

Firstly, we are going to have a look at how we design primers for specific regions of a DNA sequence. Primer design can be done manually or by using an appropriate software tool.

1. Click on the Primer design link in blackboard to open the Primer 3 web page.
2. Copy the sequence above and paste it into the first box on the web page.
3. Change the product size to min 1800, opt 2000 and max 2200.
4. Change the primer size max to 24
5. In Included region put 1, 2200 (this is a specific region with which we want to design our primers within our sequence, 1 being the first codon and 2200 being the base region upto where we want our primers)

.....
2761 TCCAACCCCTGCCTCTCCATGGCCTTGCTCAACTCAGCCACAACATCCTCACCAACCTGA
.....
2821 TTAGGGAGTAACCAAATAACAAATTAATAAATAAAGTAATTGTGGTTATTATGGGATGATT
.....
2881 GAAAAATTGGCTCAGAGACCATCAACCGAGAACTTAATATTCTTAAGTATCACAAATAT
.....
2941 ATTCTTCCTTACCAAGCTCTACATTTTCATCGTTGTGATAAACTCAGATAACATCAACCCA
.....
3001 GAAACATACTACTCTTACTGCATCATAAAAAATGTGCTTCCCTAACCAAGCTTTACATTTCA
.....
3061 TCATTGAGATAAAGAAATCAGTTTGACCATAATGTAATGAGTGATAACGAAGTATCAGTG
.....
3121 AACTAAATGAATCAACATGAAATAATATTTAATAACATTTTCATCATTGAGATAAAGAAAT
.....
3181 CAGTTTGACCATAATGCAATGACTGATTGAGAAGTATCAGAGAACTAACTGAGTCAAAAT
.....
3241 GAAATAATATTTAATAGCCACACGCCACACCACTATTAAAGTTCAACACTTGTGGCATA
.....
3301 TACAAATATACAATGGTTCAGAAGGCAAACCTCTTTTAGCCCTCCATGCCATCTAGAAAT
.....
3361 GACTAAACATCTCTATTATATAGTTAACTAAATTAATAATGAAAGTGATTGACGAAGAG
.....
3421 ATGACAAGTAGTACAATAGAAAAAATGTCATAAGTGCTAAGAGGCAAATCCTTTTTTTCAT
.....
3481 TTCTCTCCTTCAAGGTCAGACATTATCTTAGATGGTTTGGCTAAGAAAAGGGTGTGTTAA
.....
3541 ATCTGCTCTACTATACCAAAGAGTGATCAAGGACAAGAACCGAGTCCCTTGCCATATCA
.....
3601 AATAAAGGTTAACCACCAGAACTTGCCATATCAATAAAAAATACACTTGAAGCCATGTCA
.....
3661 ACTAATGTCCTAGGAGTTTGTGGACATAATCTGCATAGTGTCTGAATTTACCGTGCCG
.....
3721 TTGATCGAAAAACCTTTGTCCACTTGATGAGCGTTCCAGATGATATCGACAATTGGTTC
.....
3781 ACTGAAAAGGAAAAGGTGAAACCCAGCTCTCTCTGCCTCCCCTCTGGGAGGTGGAAATCG
.....
3841 TTGCCTTCGGTATCCACAAAATTTGCCAATGCAGCAGCAATGAAATCAAACAGTTCCTGT
.....
3901 ATCAATAGTTATTAAAGATACATTAGGAACCACGCGCTAAATTTGGCAACTCCAGGAAA
.....
3961 CTTGAGAATCAAACAGCAGGAACTCACCATGGAAGTCCCAACCATGAGGTGAGGTGGGA
.....
4021 TAGAGACTTCCTCGGACTGTGCTTGGCAACACGTTTCTCCCTTCTCGGAGTTGGACTC
.....
4081 GGAGTACACGGAAGTTGGTTCCTCCAAGATCCAGCGCATAAAACAGCCCATGCTCGTCTC
.....
4141 TGCAGACATAGCACCCAGATGATTAAACACAAAAGTTCAAGAGTCAAGAAAAAAATGCA
.....
4201 TGTTTATCAGTGAATTGCACATAATAGTCTGAACGCAACAAAATGTTAGATTTTAAGAAC
.....

4261 TTGAGTTTCAGTGAATGATGCTCAACATGTTAAAGCATTATATACTATCTGTTATAATF
.....
4321 CGTGACCAATGTTTTCTCAACATGAGTTCGATCTGAAATTCCTAGTGTATAAGAGGATG
.....
4381 ATCCTAGAAAACTAAGCAATTCACAGGTTATACGATTAGGTCGAGTAATTGATATAACT
.....
4441 CAATAGAGAAATGCTGACACAAAACCATAAAGATGTATCAACGTTTCTAAAGAATTAAGA
.....
4501 GAATTGAGCTTATAGGAGTAAATCTGAGATAACTACGCATGTCATTAATCGAAAACTAA
.....
4561 GCACTTGACCGATTATATCTAGAGATGATTAGGTGGACTGGCAGATATAACTGAACAAAC
.....
4621 AAATTATTGAAAGAAACGATAAAGACATATGAACTTTTCTACAGAATTCACAGTATAGGT
.....
4681 GTAAACATGCGCTAACTATCAATGCCATTAATTGGACAGTACATGTACATCTCACTAACG
.....
4741 GGCTATCAAATAGGCTCGCCCCACCAGGCACTCATCGCAATCTCATCATGTCACTCCAAG
.....
4801 ACTGGTGACATTCTCATTTGACCCATATCCTAAATCATCAAGGTTGTTTGGAACTACCA
.....
4861 CGTGATCTGCAAGCATGGGATTTGGCAGGGCGGAATTGGTAGGTAAAAGTATCCTATCCT
.....
4921 GGCCCATGATCCATCTAGAATCGGCAGAAGGGTCGATCTTCGCAATAAGATCCGGGCAT
.....
4981 TTCGCCGCGAGGGCGGTCAATCCTAATCAAGAGCGGGAGAGAAGAATTGCTGACGCTTCC
.....
5041 GCGATCAGGACTCAGAATTCGTCCAGGCGCGACGACAGTCCCAACAACAGACGGCGTCT
.....
5101 TTGGGATCACAACAACAAGTCAACCGAACCGCAGAAAAGGAAAGGAAAGGGAAAAAAG
.....
5161 GAAAGACGGGATCGGATCGGATCAGATCGGACAAGAATGGGATCTTTGTTACCCGGTGGG
.....
5221 GAGGTTGTCGACGTATGAGATGAGCATCTTGAGCTGTGAGTGGATGTCCCCGCGCAGTCC
.....
5281 GCGCTCCATCTCGGCGACGAGCGCGTCCGAGATGCTCCTGAGCAGGGCGGTGGGCGTGA
.....
5341 GAGCGCGCGCTCGACCTCCTCGATCAGTCCGCCACCTTGCGCTTCGCTCGGCCTCGGC
.....
5401 GCGCGCGTGCAGCTCCGCGCCCGCTTCTGGCGGCGCCGCCAGCAGACCGCCGTGCC
.....
5461 CACCGCCGCGCACACCACCACCGCGTGCCACCAGCGCCGCTTCCCCATCGCTGAGAT
.....
5521 CAGCCTCTCTCCGCCAGCGAACGAGCGATCGACCGGCAAGAATCGCAGAGGAATTCCGC
.....
5581 TCTGGATCCGAGGGAGTGAAGGAAGGGATGGTTCGCCCCGGCGGTAGGAGAGGGACTGG
.....
5641 CCCTCGGCGTTTTAACCGGTAACTTTAACTGGCCCTTGCTTTGCGGCATTTATGGATCC
.....
5701 ACCTATAAATACACGAACGGCCGGGATTTGTCCGGGGACACGTGGACGGACGGGATCGT
.....
5761 GGCTTGGAGAAGTGGATCTGGATTTTTTTGGATTATTAGGGAACGGATTTTCTGCTGTG
.....

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5821 GAAATATTACTAGACGGATTTACTCCCAGATATACTTTACGGTAAGTACCTCCCGTGTA
.....
5881 TTACCTACGGAGTATATGATTTGGCGAGTAGATCCTGAATTAGAGCGCTGACTTGTGGCG
.....
5941 GCGGCCCGCGTCCCAGTGGG
.....

```

```

KEYS (in order of precedence):
..... vector sequence
>>>>> left primer
<<<<<< right primer

```

ADDITIONAL OLIGOS

	<u>start</u>	<u>len</u>	<u>tm</u>	<u>gc%</u>	<u>any</u>	<u>3'</u>	<u>seq</u>
1 LEFT PRIMER	25	19	60.73	57.89	4.00	2.00	GCAGCTTCGACATCGTCTG
RIGHT PRIMER	2020	20	60.08	40.00	7.00	0.00	ATGATGCTGCATTGTTTGGA
PRODUCT SIZE: 1996, PAIR ANY COMPL: 6.00, PAIR 3' COMPL: 0.00							
2 LEFT PRIMER	20	18	59.94	55.56	4.00	2.00	CAAGGGCAGCTTCGACAT
RIGHT PRIMER	2020	20	60.08	40.00	7.00	0.00	ATGATGCTGCATTGTTTGGA
PRODUCT SIZE: 2001, PAIR ANY COMPL: 6.00, PAIR 3' COMPL: 1.00							
3 LEFT PRIMER	25	19	60.73	57.89	4.00	2.00	GCAGCTTCGACATCGTCTG
RIGHT PRIMER	2018	20	59.81	45.00	7.00	0.00	GATGCTGCATTGTTTGGA
PRODUCT SIZE: 1994, PAIR ANY COMPL: 6.00, PAIR 3' COMPL: 2.00							
4 LEFT PRIMER	20	18	59.94	55.56	4.00	2.00	CAAGGGCAGCTTCGACAT
RIGHT PRIMER	2018	20	59.81	45.00	7.00	0.00	GATGCTGCATTGTTTGGA
PRODUCT SIZE: 1999, PAIR ANY COMPL: 6.00, PAIR 3' COMPL: 1.00							

Statistics

	con	too	in	in	no	tm	tm	high	high	high	high	high	
	sid	many	tar	excl	bad	GC	too	too	any	3'	poly	end	
	ered	Ns	get	reg	GC%	clamp	low	high	compl	compl	X	stab	ok
Left	2669	0	0	0	10	0	1625	322	39	50	0	32	591
Right	2663	0	0	0	20	0	1786	185	1	16	0	45	610

Pair Stats:

```

considered 2225, unacceptable product size 1767, high end compl 54, ok 404
primer3 release 1.1.4

```

The arrows below the lines of sequence show the location of the primers. From this output we can see the top primers picked by Primer 3 and some alternatives below the sequence output. This way we can choose which primers we deem the most suitable depending on the criteria.

VALIDATE OUR PRIMERS

Next we need to validate our primers to check that they will amplify a region of sequence specific to what we need.

1. **Click on the NCBI Primer BLAST link on Blackboard**
2. **Copy and paste the left primer and right primer output from primer 3 into the forward primer and reverse primer parameter boxes respectively.**
3. **Change the PCR product size to 1000 min and 2200 max**
4. **Change the Organism box in primer pair specificity checking parameters from homo sapiens to plants.**
5. **Click show results in a new window and select GET PRIMERS**

We should now see a list of products that the primers are hitting within the database and we can check that these are hitting sensible results. This enables us to check the selectivity of the

primers we are designing and whether we need to go back and redesign some more appropriate primers.

2. DNA Sequence Mutations

In this module we have determined the presence of a nematode drug resistant strain and a drug susceptible strain. From the drug screening assay we looked at in workshop 1 we are able to identify and observe the phenotype using microscopy. To further analyse the presence of any drug resistant strains and the drug susceptible strains, we can extract DNA from the worms and used PCR to amplify a region of DNA in order to determine whether a potential sequence mutation may be present in our nematode strains.

In this section of the workshop we will align the DNA sequences from the nematode stains and identify any sequence mutations. We will then BLAST the sequences using the NCBI database

The DNA sequences below are from our laboratory samples:

Susceptible strain

```
tagtgaagttagtttagataatgaaaatgtataagaaaaccctataaaatgtttcgactttttgttcaatgtttatta
aaaagtgtctatttctgagttctgaacttccacaatttgaaagtcgacaaaaaatgatttctatattttgatcaga
ctaagggcataattatftaagcattatagatcttcaaacacgcaataatactatatttctcaaaaagacaaaatatttcg
agaagttggcaagtcgaaatttcaaagttccctaaactaatggaatgtagtacattttcatattcttcgttttgctgat
ttctttaatttccacaacatatttttaaaagtaaatacctcagatctccagattcttcatatctttcataagccaa
tctttgatctgtccatttctctgaaatgtgaattgagcactgtattccatatttacgcatcaattttgatattgatc
gaagataaatgtttactgtcactagaacgggtccaccggtatctgatagtttagttttgaattaattctgcataatag
aagtttaaattgtattttgccaaggttgcaaattttcaagggtcaaaaatctttgaaatactttttgggattatg
caataagtggttttcaagtgcaaaactcaatttctacagaaaaactcctgaacctgccgattcagcaaagtaaatt
tcaagaacctattttcacagttttcaaaatctagattaattcatataggttcggtgcttagtaattctcggtgagac
ccatctgaatcgagttgcttgctataggtcagcgtttcagaaaaatgcaaaacaactataactattgttttcattag
ttaacttggttctgaattcaattccaataatttctgaatttaaaaactaaaaatactcacctggccacgtggcattca
ttccacgtggtctaacttccaatcatagtctttcaatatt
```

Resistant strain

```
tagtgaagttagtttagataatgaaaatgtataagaaaaccctataaaatgtttcgactttttgttcaatgtttatta
aaaagtgtctatttctgagttctgaacttccacaatttgaaagtcgacaaaaaatgatttctatattttgatcaga
ctaagggcataattatftaagcattatagatcttcaaacacgcaataatactatatttctcaaaaagacaaaatatttcg
agaagttggcaagtcgaaatttcaaagttccctaaactaatggaatgtagtacattttcatattcttcgttttgctgat
ttctttaatttccacaacatatttttaaaagtaaatacctcagatctccagattcttcatatctttcataagccaa
tctttgatctgtccatttctctgaaatgtgaattgagcactgtattccatatttacgcatcaattttgatattgatc
gaagataaatgtttactgtcttagaacgggtccaccggtatctgatagtttagttttgaattaattctgcataatag
aagtttaaattgtattttgccaaggttgcaaattttcaagggtcaaaaatctttgaaatactttttgggattatg
caataagtggttttcaagtgcaaaactcaatttctacagaaaaactcctgaacctgccgattcagcaaagtaaatt
tcaagaacctattttcacagttttcaaaatctagattaattcatataggttcggtgcttagtaattctcggtgagac
ccatctgaatcgagttgcttgctataggtcagcgtttcagaaaaatgcaaaacaactataactattgttttcattag
ttaacttggttctgaattcaattccaataatttctgaatttaaaaactaaaaatactcacctggccacgtggcattca
ttccacgtggtctaacttccaatcatagtctttcaatatt
```


We can conduct a sequence alignment on our sequences to see if we can observe any mutations occurring between our susceptible and resistant strains.

1. **Click on the NCBI alignment link in Blackboard.**
2. **Copy and paste the first sequence into the box at the top of the page and the second sequence in the box below it on the NCBI alignment webpage.**
3. **Make sure highly similar sequences (megablast) is selected**
4. **Click show results in a new window**
5. **Click BLAST**

The results showing the alignment of the two sequences will be shown in a new window and we can now look to see if there are any mutations within our sequence. The sequences should be very similar as they are from the same species of nematode.

Click on Graphics this will bring up a new window (here we will be able to see if any sequence changes are highlighted). Here we can see exactly what and where the mutation is occurring.

Click on the button with the highlighted mutation in the middle of the page (zoom to sequence). We can then see the two sequences aligned and the highlighted mutation in red.

You can use the pan left and right arrows to look along the sequence for any further changes within the two alignments.

Sequence BLAST

We can also BLAST our sequence to see which organism it hits within the NCBI database.

1. **Click on NCBI BLAST in Blackboard**
2. **Click on nucleotide BLAST**
3. **Make sure align sequences is unchecked**
4. **Copy and paste the top sequence in the box at the top of the webpage.**
5. **In the Organism box type Metazoa**
6. **Click results in new window**
7. **Click BLAST**